eurofins

# RECOMMENDED PRESERVATION AND CONTAINER GUIDE

- SOLID
- BIOTA
- AIR
- PRODUCT

**At Eurofins Environment Testing, we are committed to the highest standards of sample integrity and analytical reliability. From collection through to laboratory receipt, every step in the sample handling process plays a critical role in ensuring accurate and defensible results. To support this, we respectfully ask that you observe the following best practices when preparing and submitting samples.**

## SAMPLE COLLECTION CONSIDERATIONS

Care must be taken during sampling to prevent loss or cross-contamination of preservation reagents. Any spillage or inadvertent transfer of preservatives between containers may alter the chemical composition of the sample and compromise the validity of results, particularly for multi-parameter testing. Many analytes are susceptible to degradation, volatilization, or chemical transformation when exposed to air, light, or temperature fluctuations. Where applicable, appropriate field filtration, immediate sealing, and light protection should be implemented to maintain sample stability.

## COOLING AND TRANSPORTATION REQUIREMENTS

To ensure the stability of sensitive analytes, all samples should be chilled to ≤6°C immediately after collection and maintained at this temperature throughout transportation to the laboratory. Microbiological samples should never be frozen and should ideally be kept between 3 and 5°C. Upon arrival, the surface temperature of each sample is measured using a calibrated infrared thermometer, recorded on the Chain of Custody (COC) form, and logged into our Laboratory Information Management System (ELVIS). Please note: Temperature blanks (control bottles for verifying temperature conditions during transport) are generally not required unless explicitly requested, such as in specific microbiological analyses of drinking water.

## CONTAINER SELECTION, PRESERVATION, AND HOLDING TIME COMPLIANCE

The preservation of analyte integrity requires strict adherence to specified protocols regarding container type, preservation method (e.g., acidification, cooling), and maximum holding times, which are calculated from the moment of sample collection. Eurofins provides detailed guidance on container selection, preservative requirements, sample volume, and holding time limits for each analytical parameter. For specific test groups, such as semi-volatiles, organochlorine pesticides, or asbestos, mandatory requirements may include sample sealing, light protection, or use of specialised containers to ensure analyte stability.

## SUBMISSION TIMING AND LOGISTICS

To support timely and valid analysis, we recommend that samples arrive at the laboratory with at least 50% of their allowable holding time remaining. This buffer helps ensure that results remain scientifically defensible and compliant with regulatory standards. For parameters with short holding times, we kindly ask that samples not be submitted late on Fridays or immediately before weekends, unless prior arrangements have been made with the laboratory.

## SOLID SAMPLES (SOIL, SEDIMENT, BIOSOLID, SOLID WASTES)

| Parameter | Container Type | Lab Analysis Portion (1) | Label | Eurofins Preferred Preservation | Recommended Holding Time (5) |
|---|---|---|---|---|---|
| Acid Sulfate Soil (ASS) | | | | | |
| pH Field Screen (pHf and pHfox) | P | 50 g | ● | Freeze ≤-20°C, Exclude air | 24 hours # |
| Chromium Reducible Sulfur Suite | P | 50 g | ● | Freeze ≤-20°C, Exclude air | 24 hours # |
| SPOCAS | P | 50 g | ● | Freeze ≤-20°C, Exclude air | 24 hours # |
| Alcohols (Ethanol, Methanol) | G | 20 g | ● | Cool ≤6°C, Zero headspace | 14 days * |
| Asbestos | | | | | |
| Asbestos (AS4964/AS5370) – Bulk | PB / P | 2 cm x 2 cm (minimum) | ● | None, Separate sample required | indefinite |
| Asbestos (AS4964/AS5370) – Mulch | PB / P | 1000 g (6) | ● | None, Separate sample required | indefinite |
| Asbestos (AS4964/AS5370) – Soil | PB / P | 50 g (6) | ● | None, Separate sample required | indefinite |
| Asbestos (NEPM / WA Guidelines) – Soil | PB / P | 500 g (6) | ● | None, Separate sample required | indefinite |
| Acid Volatile Sulfide (AVS) / AVS-SEM | PB | 100 g | ● | Freeze ≤-20°C, Exclude air | 24 hours # |
| Bromide | G | 10 g | ● | Cool ≤6°C | 28 days |
| Bulk Density | G | 500 g | ● | None, Separate sample required | 14 days * |
| Carbon, Total Organic (TOC) | G | 20 g | ● | Cool ≤6°C | 28 days |
| Cation Exchange Capacity, Exchangeable Sodium Percentage, Exchangeable Cations | G | 50 g | ● | Cool ≤6°C | 28 days |
| Cations (Na, Mg, K, Ca) | G | 50 g | ● | Cool ≤6°C | 6 months |
| Chloride | G | 10 g | ● | Cool ≤6°C | 28 days |
| Clay Content | G | 100 g | ● | Cool ≤6°C | 14 days |

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(2) Soil samples for cyanide analysis (in cores or jars) must be wrapped in dark plastic and kept cool at < 6 °C without further treatment.
(3) Samples may need to be extracted as soon as possible if NFDHA (Nonafluoro-3,6-dioxaheptanoic acid) is an important analyte.
(4) Contact laboratory for specific advice and specially cleaned containers in accordance with US EPA Method 1633
(5) References: US EPA SW846, NEPM, NAGD, EPA VIC, ISO5667, Queensland Acid Sulfate Soils Management Advisory Committee (QASSMAC), US EPA Methods 821, 23, 533 & 1633A
(6) Whole bottle extraction is required.
(7) Do not fill the bottle past the shoulder, to allow room for expansion during frozen storage.
(8) No published holding times exist for this sampling method.
* Extract within 14 days and analyse within 40 days (Eurofins preference is to extract within 7 days for volatiles where sufficient time available).
** Eurofins preference is to analyse as soon as possible
*** Shelf life in matrix (sand) is low if not frozen, therefore freeze if storing or return to lab with samples immediately
# Where acidity (hence liming rates) is captured in actual and potential acidity, analysis within one week should be satisfactory. Samples can be dried at 80°C to extend holding time
## A minimum of 6 kg of sample is required on an air-dried basis. If the sample is not air-dried, a larger quantity will be required to ensure sufficient material is available for analysis.
### Except Metals / OCP / PCB 28 days
^ Excepting vinyl chloride, styrene or 2-chloroethyl vinyl ether, for which the holding time is 7 days with the same preservation

α If whole fish, field-filleted fish, or other seafood samples are frozen within 48 hours of the time of collection (i.e., either in the field or as soon as the samples are received at the laboratory or other storage facility), the holding time for tissue samples does not begin at the time of collection, but when further processing for analysis begins (e.g., lab-filleting, grinding, or removal of specific tissues for analysis).

### CONTAINERS

| | |
|---|---|
| PP | Plastic (polypropylene, No Teflon liner) |
| G | Glass (Teflon lined lid) |
| T | Tube |
| C | Canister |
| PB | Zip-Lock Plastic Bag |
| PET/S | Plastic Sterile |
| FH | Filter holder with cowl (Mixed Cellulose Ester Filter) |

| Parameter | Container Type | Lab Analysis Portion (1) | Label | Eurofins Preferred Preservation | Recommended Holding Time (5) |
|---|---|---|---|---|---|
| Coal Tar (Qualitative) | G | 150 g | | Cool ≤6°C | N/A |
| Coal Tart (Quantitative) | G | 10 g | | Cool ≤6°C | N/A |
| Conductivity (EC) | G | 20 g | | Cool ≤6°C | 7 days |
| Cyanide (2) | G | 20 g | | Cool ≤6°C | 14 days |
| Dioxins & Furans / Dioxin-like PCBs | G | 50 g | | Cool ≤6°C | 12 months |
| Ethylenediaminetetraacetic acid (EDTA) & Nitrilotriacetic acid (NTA) | G | 100 g | | Cool ≤6°C | 7 days |
| Explosives | G | 50 g | | Cool ≤6°C | 14 days * |
| Extractable Organofluorine (EOF) | PP / HDPE | 10 g | | Cool ≤6°C, No Teflon (PTFE) liner | 28 days |
| Fluoride | G | 10 g | | Cool ≤6°C | 28 days |
| Foreign Materials (RMS NSW T276) | PB | 6 kg ## | | None, Separate sample required | 14 days |
| Grain Size | G | 250 g | | Cool ≤6°C | 28 days |
| Hexavalent Chromium ($Cr^{6+}$) | G | 20 g | | Cool ≤6°C | 28 days |
| Iodide | G | 20 g | | Cool ≤6°C | 28 days |
| Loss on Ignition | G | 50 g | | Cool ≤6°C | 7 days |
| Metals / Metalloids | | | | | |
| Metals (Mercury) | G | 20 g | | Cool ≤6°C | 28 days |
| Metals (Other) | G | 20 g | | Cool ≤6°C | 6 months |

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|---|---|---|---|---|---|
| Microbiological Analysis | | | | | |
| Micro – E. Coli / Coliforms | PET/S | 30 g | ● | Sterile, Cool ≤6°C | 48 hours |
| Micro – Salmonella | PET/S | 50-250 g | ● | Sterile, Cool ≤6°C | 48 hours |
| Micro – Other Microbiological Test | PET/S | 30 g | ● | Sterile, Cool ≤6°C | 48 hours |
| Spore-Traps | Cassette | 1 | N/A | <40°C | Not defined (8) |
| Tape-Lifts | Tape-Lift | 2 x 2 cm | N/A | In sealed slide mailer | Not defined (8) |
| Swabs | Swab | 1 | ● | Sterile, Cool 2 - 5°C | 48 hours |
| Moisture Content | G | 20 g | ● | Cool ≤6°C | 14 days |
| Microplastics | G | 100 g | ● | Metal Lids | N/A(8) |
| Net Acid Generation (NAG) & Net Acid Production Potential (NAPP) | PB / P | 50 g | ● | Freeze ≤-20ºC, Exclude air | 7 days |
| Nitrogen | | | | | |
| Nitrogen – Ammonia | G | 10 g | ● | Cool ≤6°C | 28 days |
| Nitrogen – Nitrite | G | 10 g | ● | Cool ≤6°C | 28 days |
| Nitrogen – NOx | G | 10 g | ● | Cool ≤6°C | 28 days |
| Nitrogen – TKN | G | 10 g | ● | Cool ≤6°C | 28 days |
| Nitrogen – Total N | G | 10 g | ● | Cool ≤6°C | 28 days |
| Organophosphorus Pesticides (OPP) | G | 20 g | ● | Cool ≤6°C | 14 days * |

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|---|---|---|---|---|---|
| Per- and Polyfluoroalkyl Substances (PFAS) | HDPE | 10 g | | Cool ≤6°C, No Teflon (PTFE) liner | 90 days |
| Per- and Polyfluoroalkyl Substances (PFAS) | HDPE | 10 g | | Freeze ≤-20°C, No Teflon (PTFE) liner (7) | 90 days (3) |
| pH | G | 20 g | | Cool ≤6°C | 7 days ** |
| Phenolics (Total) | G | 20 g | | Cool ≤6°C | 14 days * |
| Phenoxy Acid Herbicides | G | 20 g | | Cool ≤6°C | 14 days * |
| Phosphorous (Total) | G | 10 g | | Cool ≤6°C | 28 days |
| Polybrominated Diphenyl Ethers (PBDE) | G | 50 g | | Cool ≤6°C | 12 months |
| Polychlorinated Biphenyls (PCB) Aroclors | G | 50 g | | Cool ≤6°C | 28 days |
| Semi-Volatile Organic Compounds (SVOC) | G | 20 g | | Cool ≤6°C | 14 days * |
| Organochlorine Pesticides (OCP) | G | 20 g | | Cool ≤6°C | 14 days * |
| Phenols – Speciated | G | 20 g | | Cool ≤6°C | 14 days * |
| Phthalate Esters | G | 20 g | | Cool ≤6°C | 14 days * |
| Polycyclic Aromatic Hydrocarbons (PAH) | G | 20 g | | Cool ≤6°C | 14 days * |
| Semi-Volatile Chlorinated Hydrocarbons | G | 20 g | | Cool ≤6°C | 14 days * |
| Semi-Volatile Organic Compounds – Other | G | 20 g | | Cool ≤6°C | 14 days * |
| Sodium Adsorption Ratio (SAR) | G | 50 g | | Cool ≤6°C | 5 days |
| Sulfate | G | 10 g | | Cool ≤6°C | 28 days |
| Total Recoverable Hydrocarbons (TRH) | G | 20 g | | Cool ≤6°C, Zero Headspace | 14 days * |

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|---|---|---|---|---|---|
| Total Recoverable Hydrocarbons (TRH) with Silica Gel Clean-up | G | 20 g | | Cool ≤6°C, Zero Headspace | 14 days * |
| Tributyltin (TBT) | G | 50 g | | Cool ≤6°C | 14 days |
| Volatile Organic Compounds (VOC) | G | 20 g | | Cool ≤6°C, Zero Headspace | 14 days * ^ |
| BTEXN | G | 20 g | | Cool ≤6°C, Zero Headspace | 14 days * |
| Halogenated Volatile Organics (HALVOL) | G | 20 g | | Cool ≤6°C, Zero Headspace | 14 days * ^ |
| Monocyclic Aromatic Hydrocarbons | G | 20 g | | Cool ≤6°C, Zero Headspace | 14 days * |
| Trihalomethanes (THM) | G | 20 g | | Cool ≤6°C, Zero Headspace | 14 days * |
| Volatile Organic Compounds (VOC) – Other | G | 20 g | | Cool ≤6°C, Zero Headspace | 14 days * |
| TRH C6-C10 / BTEX for Trip Spikes / Blanks – Prepared in the Lab | G | Full jar | | Freeze ≤-10ºC (7) | 14 days *** |

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## BIOTA SAMPLES

| Parameter | Container Type | Lab Analysis Portion (1) | Label | Eurofins Preferred Preservation | Recommended Holding Time (5) |
|---|---|---|---|---|---|
| Per- and Polyfluoroalkyl Substances (PFAS) | | | | | |
| Fish (Fillets or Whole) | PP / PB (4) | 25 g | | Freeze ≤-20°C, No Teflon (PTFE) liner (7) | 90 days (3, α) |
| Crustacea (Whole, White flesh or hepatopancreas) | PP / PB (4) | 25 g | | Freeze ≤-20°C, No Teflon (PTFE) liner (7) | 90 days (3, α) |
| Shell fish (Whole or Edible flesh) | PP / PB (4) | 25 g | | Freeze ≤-20°C, No Teflon (PTFE) liner (7) | 90 days (3, α) |
| Milk［Bovine, Ovine, Caprine］ | PP | 500 mL | | Cool ≤6°C (DO NOT FREEZE), No Teflon (PTFE) liner | 28 days |
| Cheese［Bovine, Ovine, Caprine］ | PP / PB (4) | 250 g | | Cool ≤6°C (DO NOT FREEZE), No Teflon (PTFE) liner | 28 days |
| Eggs［Chicken］ (White, Yolk or Whole) | PP / PB (4) | 6 each | | Cool ≤6°C (DO NOT FREEZE), No Teflon (PTFE) liner | 28 days |
| Vegetables［Lettuce, Cabbage］ | PP / PB (4) | 250 g | | Cool ≤6°C (DO NOT FREEZE), No Teflon (PTFE) liner | 28 days |
| Fruit［Avocado, Tomato, Olive, Mango］ | PP / PB (4) | 250 g | | Cool ≤6°C (DO NOT FREEZE), No Teflon (PTFE) liner | 28 days |
| Crops［Sorghum, Wheat, Corn, Canola］ | PP / PB (4) | 250 g | | Cool ≤6°C (DO NOT FREEZE), No Teflon (PTFE) liner | 28 days |
| Microplastics | | | | | |
| Biota | G | 100 g | | Cool ≤6°C, Metal Lid | Not defined (8) |

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|---|---|---|---|---|---|
| Volatile Organic Compounds (VOC) and/or CRC Care TR23 | | | | | |
| Summa Canister | T | 1 L or 6 L | | Ambient Temperature | 30 days |
| Low Uptake Waterloo Membrane Sampler (LU-WMS™) | T | 1 | | Cool ≤6°C | 30 /28 days |
| US EPA Method 325B Passive Sampler | T | 1 | | Cool ≤6°C | 30 days |
| Radiello® Passive Sampler RAD130 | T | 1 | | Cool ≤6°C | 30 /28 days |
| Radiello® Passive Sampler RAD145 | T | 1 | | Cool ≤6°C | 30 days |
| Dioxins & Furans / Dioxin-like PCBs | | | | | |
| XAD-2 Resin | T | 1 | | Cool ≤6°C | 30 days |
| Asbestos | FH | 1 | | Ambient Temperature | Not defined (8) |

## PRODUCT SAMPLES

| Parameter | Container Type | Lab Analysis Portion (2) | Label | Eurofins Preferred Preservation | Recommended Holding Time (5) |
|---|---|---|---|---|---|
| Total Fluorine (TF) | | | | | |
| Cosmetics | PP / PB (4) | 25 g | | Cool ≤6°C, No Teflon (PTFE) liner | 28 days |
| Food Packaging | PP / PB (4) | 2 cm x 2 cm | | Cool ≤6°C, No Teflon (PTFE) liner | 28 days |
| Textiles | PP / PB (4) | 2 cm x 2 cm | | Cool ≤6°C, No Teflon (PTFE) liner | 28 days |

### NOTES

(1) The specified sample volume is based on the amount sufficient for a single test, not the capacity of the sample containers provided by Eurofins. We recommend that you provide additional sample on the 1st, 11th, 21st, 31st etc sample for performance of Duplicates / Matrix Spikes. (Note however that Matrix Spike determinations are not appropriate for all tests).
(2) Soil samples for cyanide analysis (in cores or jars) must be wrapped in dark plastic and kept cool at < 6 °C without further treatment.
(3) Samples may need to be extracted as soon as possible if NFDHA (Nonafluoro-3,6-dioxaheptanoic acid) is an important analyte.
(4) Contact laboratory for specific advice and specially cleaned containers in accordance with US EPA Method 1633
(5) References: US EPA SW846, NEPM, NAGD, EPA VIC, ISO5667, Queensland Acid Sulfate Soils Management Advisory Committee (QASSMAC), US EPA Methods 821, 23, 533 & 1633A
(6) Whole bottle extraction is required.
(7) Do not fill the bottle past the shoulder, to allow room for expansion during frozen storage.
(8) No published holding times exist for this sampling method.
* Extract within 14 days and analyse within 40 days (Eurofins preference is to extract within 7 days for volatiles where sufficient time available).
** Eurofins preference is to analyse as soon as possible
*** Shelf life in matrix (sand) is low if not frozen, therefore freeze if storing or return to lab with samples immediately
# Where acidity (hence liming rates) is captured in actual and potential acidity, analysis within one week should be satisfactory. Samples can be dried at 80°C to extend holding time
## A minimum of 6 kg of sample is required on an air-dried basis. If the sample is not air-dried, a larger quantity will be required to ensure sufficient material is available for analysis.
### Except Metals / OCP / PCB 28 days
^ Excepting vinyl chloride, styrene or 2-chloroethyl vinyl ether, for which the holding time is 7 days with the same preservation

α If whole fish, field-filleted fish, or other seafood samples are frozen within 48 hours of the time of collection (i.e., either in the field or as soon as the samples are received at the laboratory or other storage facility), the holding time for tissue samples does not begin at the time of collection, but when further processing for analysis begins (e.g., lab-filleting, grinding, or removal of specific tissues for analysis).

### CONTAINERS

PP Plastic (polypropylene, No Teflon liner)
G Glass (Teflon lined lid)
T Tube
C Canister
PB Zip-Lock Plastic Bag
PET/S Plastic Sterile
FH Filter holder with cowl (Mixed Cellulose Ester Filter)

eurofins

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