

- The sampler will receive a sample kit from our lab.
- WHEN SAMPLING, BRING ICE IN SEALED BAGS TO CHILL SAMPLES DURING SAMPLE COLLECTION.**
- Put on nitrile gloves. If sampling from faucet, remove the aerator and screen.
- Open the tap and let the water of the sample source run at fast flow for approximately 5 minutes.
- The sample kit will include clean, baked amber or clear borosilicate glass vials. Volumes and preservatives required per test are as follows:

TEST NAME	BOTTLES	PRESERVATIVE	HOLD TIME
Haloacetic Acids (SM6251B)	(3) 40mL amber vials	65mg NH ₄ Cl xls/vial	9-14 days
Trihalomethanes/HANS (EPA551.1)	(4) 60mL amber vials	Buffer +NH ₄ Cl/vial	14 days
Trihalomethanes (EPA524.2)	(4) 40mL amber vials	thiosulfate/vial	14 days
Chloral Hydrate (EPA551.1)	(3) 40 mL clear vials	buffer + sulfite /vial	14 days
Aldehydes (EPA556)	(2) 40 mL amber vials	CuSO ₄ + NH ₄ Cl/vial	14 days
Bromide/Bromate/Chlorite/Chlorate (EPA300.0/EPA300.1)	(1) 60 mL poly	EDA/bottle	28/14/28 days
Chlorine Residual (SM4500-CL)	(1) 250 mL amber	None	Immediate

- Use indelible ink (i.e. Sharpie pens) to clearly identify the sample bottles with the information listed below (if not already on the label).
 - Client Name - Analysis required - Preservative used
 - Sample ID - **Date and Time of collection**
- Slow water flow to thickness of a pencil (to minimize splashing) and fill bottle.
- Fill ALL 40-mL or 60mL vials, slightly overfilling them. For plastic bottles, it is okay to leave headspace. Make sure the mouth of the bottle does not come in contact with anything other than the sample water. **DO NOT RINSE OUT PRESERVATIVE.**
- Cap and invert the vials/bottles at least 5 times to mix the sample and preservative. **CAUTION:** Invert sample vial and tap it to check for trapped air bubbles. If air bubbles are detected, add additional sample.
- Store at ≤6°C but above the freezing until transported to the lab.

SAMPLE SHIPPING AND STORAGE

- If shipping samples on the same day of sampling, chill samples until ≤6°C by exchanging the wet ice used during sampling with **FRESH** wet ice.
- Pack chilled samples** in a cooler and add enough **FRESH** wet ice to take up 30-50% of the cooler (e.g. most of the remaining space) inside two large plastic bags as recommended in our **“Wet Ice Packing Instructions.”**
- Complete the Chain of Custody during sample collection. Place Kit Order and completed Chain of Custody in a Ziploc style bag in the cooler on top of packing material. The following information is required on the completed Chain of Custody.
 - **Collector’s name** - **Sample site** - **Comments about the sample** (if applicable)

- Client Name -Date and time of collection -Sample type

4. **Ship via overnight service such as FEDEX, UPS, or DHL, etc.** Maintain an environment at $\leq 6^{\circ}\text{C}$ but above the freezing point of water during transit. It is recommended that samples arrive within 48 hours of sampling, with no more than 40 hours for transit.
5. If samples are received on the same day as collection, temperature may be $>10^{\circ}\text{C}$ with evidence of cooling.
6. Maximum **HOLDING TIME FOR SAMPLES** for Chlorine Residual is **immediately** from time of collection. For other hold times, please see the above table.
7. Alternatively, cool the samples down by placing them **overnight** in a cooler with wet ice, or in a refrigerator (store chilled for at least 12 hours before packing for shipment). Maintain the cold samples until repacked in the cooler for shipment to the lab.

ADDITIONAL NOTES

- Try to collect only on a Monday, Tuesday or Wednesday and ship no later than Thursday of each week, and try to **NOT** collect samples on Friday, Saturday, or Sunday unless special arrangements have been made for the receipt of samples at the laboratory within 48-hours of collection.
- **FOR PLANTS USING CHLORINE DIOXIDE:**
When collecting samples from treatment plants employing chlorine dioxide, sections 4.6 and 8.4 of EPA Method 300.1 requires that the samples be **sparged** with an inert gas (helium, argon, nitrogen) for approximately 5 minutes prior to the addition of EDA preservative at the time of sample collection. Otherwise, chlorine dioxide will continue to form chlorite. EEA bottles must be filled **after** this purge step. For detailed sparging instructions, see **Chlorite for Plants Using Chlorine Dioxide**.
- If shipping to the laboratory with **frozen gel packs** rather than wet ice, please be sure that the gel packs have **been frozen for at least 48 hours** prior to the shipment time.