

Lancaster Laboratories

Development of a Simple LC-HRMS-based Disulfide **Mapping Assay Applied to Biopharmaceuticals**

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OVERVIEW

Purpose: To develop a simple 3-step reaction LC-HRMS-based disulfide mapping method for common biopharmaceuticals.

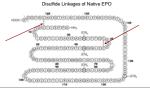
Methods: Controlled reduction, cyanylation, and

Orbitrap MS.

<u>Results: The method</u> was able to produce and identify all distinguishable modified peptides of the native structure of Recombinant Human Insulin (rH peptides representative of two possible scram products of rH Insulin.

INTRODUCTION

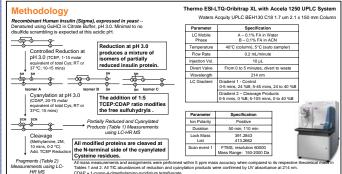
- Disulfide bond formation in proteins is an important PTM essential for the stabilization of its 3D structure.
- A simple assay is presented with no HPLC fractionation involved that can deduce the lin closely-spaced cysteine residues.
- Three proteins with different disulfide mapping complexity defined by the number of disulfide lini cysteine residues proximity, and protein size were evaluated as test proteins: Ribonuclease A and
- Rivase A was rully characterized by JI watson and colleagues. Results were published in various journal Erythropoietin has four Cysteine residues all involve in two disulfide linkages. EPO is easy to characterize using in-solution tryptic digestion in its non-reduced



since no endoproteinase can digest it to produce peptides with one disulfide linkage.



METHODOLOGY and RESULTS



Recombinant Human Insulin



Controlled Reduction and Cyanylation Products

Objective: Formation of a Mixture of Insulin Protein Isomers with 1 and 2 Disulfide Linkages Reduced and Cvanvlated

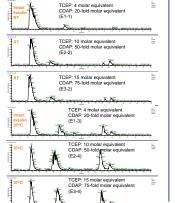
Experimental Design:

- Total Number of Cysteines in Protein
- [TCEP]_{min} = 4 molar equivalent for every Cys
- [CDAP1.... = 20 molar equivalent for every Cvs;
- [CDAP] should be minimum 5x of [TCEP]
- Both reactions performed at pH 3

	Mass-to-Charge Values, PLX							
	8.0	JR-10"	29-242	26-342	Mr mo-	M- 90*	Je- 90*	De DC
Intact Non-Reduced rH Insulin	5863.6371	5801.6444	2902.6209	1435.5530	1451,9166	1161.7347	968.2802	830 0063
Controlled Reduction P	roducts							
Two Cys - scacs of I legaler	5805.8517	3606.8590	2923.6332	1939,2249	1452,4202	1162,1376	308.6100	E30 5861
Four Cys - intact (H Insulin	5807 8663	1008.6736	2904.8405	1936,8961	1452.0030	1162,5406	368 3517	830 6730
Two Cys.: A-Chan (108 resolven)	2379:0042	2000.0015	1190 9994	794,3354	590,0034	477.0041	397.6713	341.0050
Two Cyn - B-Chart (fully	3427 6045	3429.0018	1714 8496	1143.5688	957 1094	696.5442	572.2mm	490.6768
Four Cys - A-Chair (fully reduced)	2382:0000	2363.0073	1196:0073	795.0078	596.5073	477.4575	396,0075	341,2930
Cyanylation Products								
Two Cyn - intact (HI Insulin	5855.6719	3858.6TM	2928 8433	1952,8979	1464.0253	1172,1417	114 3504	837.5319
Four Cys - Intect IHI Intel®	Asset tour	900K 7140	State most	1979.3479	1477 3045	1102 5400	100 420 1	BAR MIN
Two Cys - A-Chair (1D8 intrachen)	2430.0044	3431.0117	1216:0095	811.0088	808.5084	487.0002	404,0080	348,1506
Two Cys - B-Onen duly	3477.7047	5478.7120	1739.8597	1160,2422	870.4335	616.5452	580 6245	497 8225
Four Cys : A-Chain (hifty reduced)	2482.0404	3483.0477	1242.0275	808.3541	621.5174	497.4154	414 0007	355 5645

Incubation at 37°C Temperature Produced More Abundant Reduction and Cyanylation Products

found in the native secondary structure



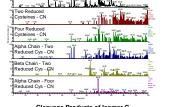
TCEP: 4 molar equivalent CDAP: 20-fold molar equivalent TCEP: 4 molar equivalent CDAP: 40-fold molar equivalent

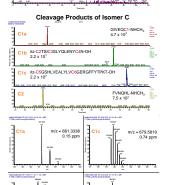
TCEP-to-CDAP Ratio was Optimal at 1:5 Minimum Identification of Reduction and Cyanylation Products Reduced Cys-CN 6.1 x 10⁷ Reduced Cvs-CN In-Chain - Two 7.7 x 10⁵ Red. Cys-CN aβ-Chain - Two 9.9 x 10⁷ Red. Cys-CN 3.4 x 10⁶ Reduced Cvs-CN

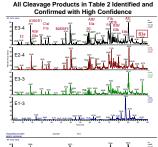
Chemical Cleavage Products

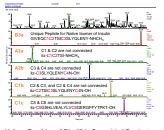
Objective: Complete N-terminal Cleavage of Cyanylated Cysteines with Minimal Side-Reaction Products

Experimental Conditions Tested: Cleavage using 1 M NH,OH, RT, 1 hour - Various forms of carbamylated insulin were the dominant product of the reaction. • Cleavage using 2 M Methyl Amine, 0-2 °C, 10 mins. 2 M CH₃NH₂ Showed Efficient Conversion of Cvanvlation Products to Truncated Peptides TIC F3-4









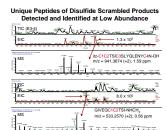


Table 2. Mass-to-charge Values of all Cleavage Products from all Partially Reduced and Cyanylated 1DB and 2DB Insulin

Sequence III. So (RH NO) - (RH 20) - (RH 40) -



CONCLUSIONS

 A simple method has been demonstrated to correctly assign disulfide linkages in a protein containing closely spaced or adjacent cysteine residue

The method is rapid, sensitive, and does not require fraction collection, but instead relies on the accurate mass measurement capabilities of the LTQ-Orbitrap

Data interpretation is time consuming because of the heavy manual calculation of the exact masses of all truncated peptides based on its elemental composition

FUTURE DIRECTIONS

nerapeutics with consecutive cysteines involved in lisulfide linkages (e.g. hinge region of mAbs). After digestion with trypsin or any endoproteinase of choice the identified non-reduced peptides containing the target regions for characterization can be fractionated or enrichment purposes. Apart from performing a systematic optimization of controlled reduction and complete cyanylation steps, the rest of the method can

When applied to proteins with unknown disulfide inkages, fractionation of the partially reduced pro somers prior to cleavage will be necessary.