

TSE Active Surveillance Information for Cattle, Sheep and Goat Fallen Stock Sampling Sites

Change notice

The following changes have been made to version 3:

- Summary Guidance for Fallen Stock Cattle, Sheep and Goats, bullet 2 updated
- Ordering, Forms, paragraph 2 updated
- Quota and Pre-Selection of Samples, paragraphs 2 and 10 updated
- Payment for Sampling, paragraph 3 updated

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Summary Guidance for Fallen Stock Cattle, Sheep and Goats

1. All fallen cattle over 48 months are required to be sampled.
2. A quota system is used for the sheep and goat fallen stock survey and it is important the quota is adhered to, to ensure good representation from all regions and holdings. Select good quality samples for testing from as wide a range of holdings as possible, and no more than three samples from any one holding per day. Changes to the quota will be emailed to you by the Animal and Plant Health Agency (APHA) on a regular basis. Only sheep and goats over 18 months should be selected for sampling.
3. It is essential that the submission forms are completed correctly, with a legible County Parish Holding (CPH) number, address, ear tag and age (dentition), particularly for sheep and goat samples. Missing information causes delay in processing the sample for testing with a possibility of not being able to be included in the survey.
4. The Fallen stock site should strive to provide cattle passport movement card for each sample submission form, as this allows APHA to record the ear tag accurately.
5. Samples submitted will be monitored to ensure that the correct sampling procedures have been undertaken: e.g. both Cerebellum and Obex are provided (sheep and goats only), submission form correctly completed with all the required information, etc. This will allow APHA to determine if the monthly sampling fee can be paid in full.
6. To ensure the traceability of sheep and goat samples, the barcode on the pot and inclusion of ear tags are vital. In the case of Compulsory Scrapie Flock Scheme (CSFS) and Scrapie Monitoring Scheme (SMS) / Scrapie Qualifying Scheme (SQS) samples the barcode is omitted, but the pots should be clearly labelled with 'CSFS' or 'SMS' / 'SQS', as appropriate. The test laboratory will apply barcodes to these samples on receipt.
7. In addition, samples such as those for the CSFS and SMS / SQS surveys should follow a clear set of instructions to distinguish them from the fallen stock samples.
8. Consumable usage is continuously monitored, and the testing laboratory have been instructed to amend orders if they are disproportionate to the number of samples being collected. Therefore, try to ensure that all consumable orders are appropriate for your sampling rate.
9. Samples should be adequately packaged for despatch, with clear collection instructions for the Courier.
10. The testing laboratory, Eurofins Forensic Services Ltd (EFS), will report the test results to APHA for active surveillance fallen stock samples only. The Scrapie monitoring scheme (SMS) and Scrapie qualifying scheme (SQS) test results will be reported to the Scotland's Rural College (SRUC).
11. Useful contact numbers:
 - consumables, test results and sample related queries - 0844 057 0110 (EFS)
 - sheep and goat quota related queries - 0345 601 4858 (CSC One Health)
 - cattle fallen stock information - 0345 601 1367 (APHA TSE Helpline)
 - CSFS queries - 0345 601 4858 (CSC One Health)

- SMS / SQS queries - 01335 320014 (National Fallen Stock Company (NFSCo)) Scotland's Rural College (SRUC) Livestock health schemes 01835 822456
- courier contact - 0800 856 2464 (Topspeed)
- Scotland's Rural College (SRUC) Veterinary Services - 0131 535 3130

Guidance Notes for TSE Fallen Stock Sampling Sites

1. This document contains useful information and reminders for cattle, sheep and goat fallen stock brainstem samplers.
2. All relevant forms and documents mentioned are attached in the appendices below.
3. The first part of this document covers cattle Transmissible Spongiform Encephalopathies (TSE) sampling and the second part includes sheep and goat. There are a number of common grounds in TSE sampling for these three species, however for ease of reading, they are written in two parts and there will be some repetition.

Overview of the Cattle Fallen Stock Project

1. The UK has been testing cattle fallen stock since 2001.
2. Cattle which die or are killed other than for human consumption are classified as 'fallen stock'. Their brainstems must be sampled and tested for TSE if they are over the following ages:
 - **over 48 months** (majority of submissions): Cattle born in the UK and all other EU Member States except for Bulgaria, and Romania
 - surveillance (BSE) cohorts and offspring killed at any age
 - over 24 months: Cattle born in Bulgaria, Romania and all non-EU Member States.

Refer to Appendix 1.

1. All cattle fallen stock samples are tested at EFS. The test used to screen samples is the IDEXX HerdChek BSE-Scrapie Antigen Test Kit. The reporting of test results has a three-day turnaround from the day of receipt at the laboratory. This test is capable of testing both fresh and autolysed (rotten) samples.
2. The APHA ABP Team will approve and monitor all sampling sites. Random checks will be carried out to ensure correct practices are followed.
3. Fallen cattle are delivered to EFS Tuesday to Saturday. A holiday schedule will be issued by APHA in advance regarding sampling and collections during bank holidays.
4. APHA will provide the following:

- sampling training - retraining as appropriate to approved fallen stock sampling sites
 - technical advice - on sample submission form completion and consumables procurement
 - free courier service to the testing laboratory
 - sample condition monitoring
5. EFS will provide the following:
- a testing service
 - supply of necessary consumable items to approved fallen stock sampling sites

Submission of Carcase for Sampling

1. Cattle owners should organise the collection and disposal of all eligible animals. Owners are responsible for payment of this service.
2. Age enquiries should be directed to the APHA TSE Helpline service.
3. If there is any doubt whether to sample and test, the general rule is 'if in doubt, send for testing'. This way animals that should be tested will not be missed. It has been agreed that all samples taken in good faith will be paid for where a sampling fee is applicable.
4. Any animal that is deemed by the owner to be unrecoverable must be reported to the APHA TSE Helpline service. Data on unrecoverable animals will be monitored by APHA. Any inaccurate reporting will be acted on.
5. **APHA TSE Helpline - age enquiries and unrecoverable reports: 0345 601 1367**

Submission Form

1. For all bovine submissions, the Fallen Stock Submission Form (FSCA2) needs to be completed. The form should have accurate information of animal details, sampling site details, and collector details. The form can be pre-populated (using the downloadable version) with sampling site etc. This could reduce the number of boxes to be completed for each carcase. The Fallen stock site should strive to provide the cattle passport movement card for each sample submission form, as this allows APHA to record the ear tag accurately.
2. From Spring 2025, the FSCA2 form will be revised to capture clinical signs presented by the submitted animal. It is the responsibility of the farmers to accurately provide this information when scheduling the carcase collection. This aligns with Targeted Active Surveillance recommendations from the World Organisation for Animal Health (WOAH).
3. From Summer 2025, changes will be implemented to data capture method for 'Reason For Death' field in the FSCA2 form. This will transition to the selection of coded options, with the aim to reduce time spent completing forms and enhancing data quality. This

change will contribute to achieve WOAH Targeted Active Surveillance recommendations.

4. **Refer to Appendix 2.**
5. Triplicate barcodes are placed onto the form in the box on the bottom right. The third barcode is placed on the copy of the submission form held on site for future reference. The cattle **movement card** should be attached to the front of the form and should **NOT** be placed over the barcode. This allows easier booking in of the two barcodes at EFS and reduces the risk of identification errors, which can delay the release of results.

Sampling

1. Cascade training by a previously approved sampler is the preferred method of training. A cascade training form should be completed and submitted to APHA.
2. **Refer to Appendix 3.**
3. Where cascade training is not possible, sampling training will be provided by APHA on request.
4. In addition to the cascade form, there is a sample assessment form which can be used for re-assessing competence if a site or person hasn't performed any sampling for over six months. Sample assessment should not be carried out on over 48 month old animal samples, as any errors will invalidate these samples for surveillance purposes. The site should sample under 48 month old animals and mark the samples 'for assessment only' on consultation with APHA, who will liaise with the testing laboratory for feedback.
5. **Refer to Appendix 4.**
6. All training (APHA or cascade) must be recorded on appropriate training record forms to provide evidence of approved samplers. Sampling sites should retain copies of these forms for their records. APHA field staff will expect to see documented evidence of training at site inspections.
7. Most damage is caused to the sample by not controlling the spinal cord end of the brainstem when entering the spoon (refer to Diagram 1 below) or entering the spoon at too shallow an angle (refer to Diagram 2 below).
8. For Bovine Spongiform Encephalopathy (BSE) testing purposes on cattle we require the Obex region of the brainstem.
9. **Refer to Appendix 9.**

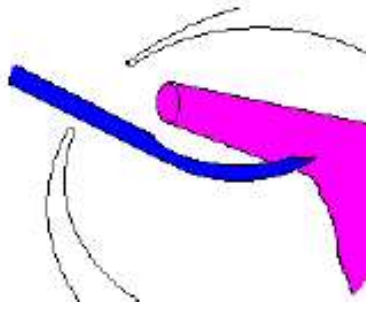


Diagram 1 - the brain is pushed into the cavity and the resulting sample is cut too short.

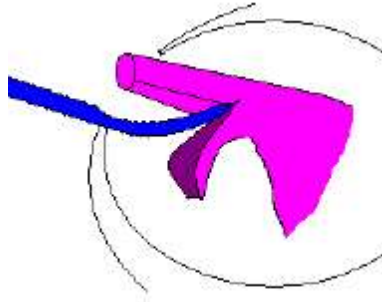


Diagram 2 - the spoon is at too shallow an angle. It slices off a piece of the brainstem. This piece often contains the target site for testing.

Autolysed (Rotten) Samples

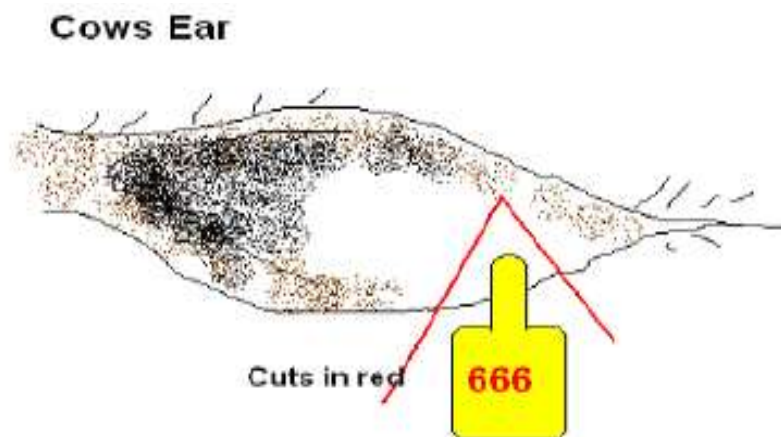
1. All brain samples can be tested despite how autolysed they are. If there is no structure to the brain that can be recognised, fill the pot about a third to half full with the brain material. Ensure the cap is fully tightened to avoid any leakage.
2. If a sample cannot be obtained, complete the paperwork and submit an empty pot with the corresponding barcode. Write 'Too Rotten' in the 'if not sampled give reason' box on the Submission Form (FSCA2).

Ear Sampling

1. APHA will require a piece of ear to be removed (a 'V' shaped notch of skin enclosing the ear tag) and placed in a clean plastic bag (refer to Diagram 3 below).
2. All individual ear samples, packed in bags, from the same day's collection should be then bagged together in another clean bag and this should be labelled with the day's date before being stored in a fridge or freezer.
3. The ear sample must be retained on site for ten working days after the matching brain sample has been sent for testing.

4. APHA will be taking random ear samples on their site visits which will be sent for DNA analysis to ensure that the site is placing the correct brain into the correctly labelled pot.
5. Also, the corresponding ear sample will be requested for any brain sample that tests positive. On receipt of this instruction the site will be requested to retain all ear samples until notified by APHA.
6. If for any reason ear tags are missing any identifying information, this information must be written down and placed in a separate clean bag with the ear tag. All untagged animals must be sampled unless there is evidence to suggest that the animal is clearly under 48 months of age.
7. If there is any mismatch of brain against ear tag, further action will be taken.

Diagram 3



Consumable Items

1. Do not over order consumable items.

Ordering

1. All ordering must be linked to the throughput of the site. We have given some guidance on the appendices. EFS encourages sites to order monthly.
2. **Refer to Appendix 5 and 6.**
3. All orders must be sent to EFS.
4. EFS will adjust all orders as they feel appropriate based on known throughput of the site and the known levels of consumables previously ordered.

Forms

1. There is also an electronic version of the Sample Submission Form (FSCA2) available for use (refer to Appendix 2). If this version is used, sites must ensure that copies are retained for their records.
2. If surveillance '**Cohorts**' or '**Home Kills**' are being sampled, correct 'Cohort' or 'Home Kills' FSCA2 submission form should be completed and reason for death box on the forms should state 'Cohort' or 'Home Kills'. These can be found in Appendix 13 (Cohort) and Appendix 14 (Home Kill). Occasionally offspring are also sampled which is required to be noted on the reason for death in FSCA2 form.

Packaging

1. There will eventually be three packaging sizes available:
 - Small Bioshield Box - (up to six samples)
 - Medium Bioshield Box - (up to ten samples)
 - Large Bioshield Box - (up to 40 samples)

Spoons

1. Disposable plastic spoons (Blue or Green) must be used for all fallen stock. Metal spoons, air or water extraction methods cannot be used.
2. Spoons should not break if used correctly, apply minimal pressure. Although EFS will normally supply blue spoons to sites, others can be supplied from time to time.

Pots

1. A common cause of complaints from the testing laboratory is when the tops of pots have not been screwed on correctly. This can lead to leakage during transit, making the packaging difficult to handle and also causing more expense as contaminated packaging has to be sent for specialist burning rather than being recycled.

Forceps

1. A pair of forceps is provided for each head. These must be treated as clinical waste.

Ear Sample Bags

1. EFS will supply a polythene bag for storage of individual ear samples. These must be stored together for that day's collection.

Barcode Labels

1. Each site will be supplied with rolls of triplicate barcodes, and each label has a prefix that relates specifically to each site. The following numbers will be sequential. One label is attached to the pot whilst its partner is attached to the submission form and the third label attached to the copy of the form held at the site for future reference.
2. Prior to using a new set of barcode labels, check the correct prefix corresponding to the sampling site is on the barcode.

Leg Tags

1. Leg tags are attached on farm to all eligible carcasses. In the event of there being a loss of ear tags this would be recorded along with farm details on the ear sample bag.

Despatch of Samples

1. APHA will be responsible for the cost of transport of samples. Currently the courier used by APHA is Topspeed.
2. Ensure appropriate numbers of collections are arranged with the courier per week based on the number of samples. Booking excessive number of collections results in additional costs to APHA.
3. Courier Topspeed contact numbers are 01565 631 840 or 0800 856 2464 and quote FALLEN_STOCK.
4. The sampling site is responsible for having the sample box packed correctly and ready for despatch.
5. **Refer to Appendix 7.**
6. Samples will need to be stored in a refrigerator before despatch.
7. Any samples not sent on a Friday should be retained in a refrigerator until the following week and dispatched Monday or Tuesday.
8. Once samples have been despatched, sites should inform EFS by emailing (bse.services@forensicsuk.eurofins.com) or faxing a Notification of Samples Sent Form (Fax: 01925 248876).
9. **Refer to Appendix 8.**
10. Bank Holiday Schedules will be issued by the APHA in advance regarding collections and delivery days. All eligible samples need to be sampled and refrigerated until the next available collection date. If in doubt regarding eligibility, sample and send them for testing. The date of birth will be validated by APHA subsequently.

Payment for Sampling

1. The cattle sampling fee will no longer be paid as of 1 April 2019, except under the circumstances outlined below for approved sampling sites.
2. The Government will pay for the TSE sampling of fallen cattle where the slaughtering is **requested by APHA** and the carcass is taken to a sampling site listed in the Carcass Collection Services Framework for the following reason:

- an animal welfare situation
 - disease situation where it is necessary to prevent the spread of disease (e.g. TB)
3. If the Contracting Body places an order that contains an animal that meets the following criteria, then a sample of that animal must be submitted for TSE testing:
- cattle aged over 48 months, or
 - cattle aged over 24 months if they were born in Bulgaria, Romania or outside the EU
4. In these instances the sampling sites are required to sample the carcass brainstem and submit this to APHA for TSE testing based on the given guidance:
- APHA will be responsible for organising the transport of the collected sample and subsequent TSE testing
 - transport is provided by Topspeed
 - testing will be provided by EFS who will also supply the consumables required for sampling and packaging
5. APHA will pay the Provider £6.25 (excl. VAT) per sample. The invoices should be submitted to tse.queries@apha.gov.uk with the following evidence:
- the order form from APHA for the carcass
 - electronic submission of animal ear tag number of the animal sampled and barcode e.g. list of ear tags on a spreadsheet
 - completion of the FSCA2 sample submission form should clearly state under 'reason for death' the animal samples was for 'TB' or 'Welfare'
 - this evidence will be confirmed against the records and if satisfied, the invoice will be paid
6. Any other invoices will be declined without payment should the above not be included.

Further Information

1. We encourage all sampling sites to look at relevant pages on the APHA website for further information.
2. Any other queries should be directed through the APHA TSE Helpline on 0345 601 1367 or tse.queries@apha.gov.uk.

Overview of the Sheep and Goat Fallen Stock Project

1. The sheep and goat fallen stock project has been in operation since 2002. The EU requirement stipulates we test fallen sheep over 18 months of age for TSE with a target sample size of 10,000 for UK. The requirement is for 9,300 from GB (93%) and 700

samples (7%) from Northern Ireland (NI). The number of sheep and goats to be sampled by individual sampling sites will be managed by APHA, based on throughput reports produced on monthly basis.

2. Owners will need to make their own arrangements and pay for the collection and disposal of all carcasses. A proportion of these animals will be eligible for sampling on a random selection basis.
3. All selected fallen sheep and goat samples are tested at EFS. The test used to screen samples is the IDEXX Herdchek BSE-Scrapie Antigen Test Kit. The reporting of test results has a three-day turnaround from the day of receipt at the laboratory. This test is capable of testing both fresh and autolysed (rotten) samples.
4. APHA will approve and monitor all sampling sites. Random checks will be carried out to ensure correct practices are followed.
5. Fallen sheep and goat samples are delivered to EFS Tuesday to Saturday. A holiday schedule will be issued by APHA in advance regarding sampling and collections during bank holidays.
6. APHA will provide the following services:
 - sampling training - retraining as appropriate to approved fallen stock sampling sites
 - technical advice - on sample submission form completion and consumables procurement
 - free courier service to the testing laboratory
 - sample condition monitoring
7. EFS will provide the following services:
 - a testing service
 - supply of necessary consumable items to approved fallen stock sampling sites

Submission of Carcasses for Sampling

1. Owners should organise with private contractors the collection and disposal of all sheep and goat fallen stock over 18 months of age. Owners are responsible for payment of this service. Age of the animal should be checked based on the numbers of permanent incisors. If there are two or more permanent incisors, then they are presumed to be over 18 months old.
2. **Refer to Appendix 11**
3. A revised Submission Form (SS2) for recording of details of farm and selected fallen stock can be found in Appendix 10. The form can be completed electronically or by hand. Pads of forms can be ordered as part of the consumables order and each pad consists of 50 forms, with a carbon copy. The top copy is sent with the sample and the yellow carbon copy needs to be retained at the sampling site.

4. Operators should ensure that handwriting is legible and the details such as CPH, animal ID and age-related information are accurate. This will be monitored by APHA, and any issues will be raised with the sites. The information provided will then be transcribed onto an IT system.

Quota and Pre-Selection of Samples

1. You will be informed of the number of sheep fallen stock samples to be taken per week in advance. The quota may change throughout the year to reflect seasonal variation, as well to ensure a good GB representation is achieved. To enable us to meet the target quota allocated to GB, it is important over-sampling is avoided, therefore we ask you stay within the quota.
2. Your collectors should pre-select eligible carcasses at farms and identify them with a numbered leg tie provided by EFS. To enable the survey to include samples from as many holdings as possible, your collectors should pre-select no more than three animals in a single collection from any holding.
3. You should then sample randomly to achieve the quotas allocated e.g. sample one in three of the pre-selected sheep and all the pre-selected goats. Ideally select good quality samples for submission.
4. If one week you fall short of the required samples, it may be possible to meet this shortfall in the subsequent weeks.
5. Barcodes are provided in triplicate. One of the barcodes is for the submission form (in the box in the middle on the right), a second barcode goes on the sample pot, and the third one is for labelling the bag containing the ear and ear tag (refer to instructions below).
6. **Refer to Appendix 10.**
7. The submission form provides space to record visible animal ID information, such as ear tag(s), tattoos or any other marking unique to the animal.
8. Sampling training will be provided by APHA. Cascade training by an approved sampler is also allowed.
9. All training (APHA or cascade) must be recorded on appropriate training record forms to provide evidence that sampling is undertaken by approved samplers. APHA expects to see documented evidence of training when sites are visited and inspected.
10. **Refer to Appendix 3.**
11. Only animals over 18 months are required to be sampled for this survey. The age can be determined by the numbers of permanent incisor teeth. Animals with two or more permanent incisors, or which are 'broken mouthed' are considered to be over 18 months. Animals suspected to be under 18 months are not eligible for sampling.
12. Any animals selected for the fallen stock survey must have intact eartags when collected.

13. Refer to Appendix 11.

14. For Scrapie testing purposes on sheep (ovine) and goat (caprine) samples we require the Obex region of the brainstem and the Cerebellum.

Example of suitable brain sample



1. Most damage is caused to the sample by not controlling the spinal cord end of the brainstem when inserting the spoon or inserting the spoon at too shallow an angle.

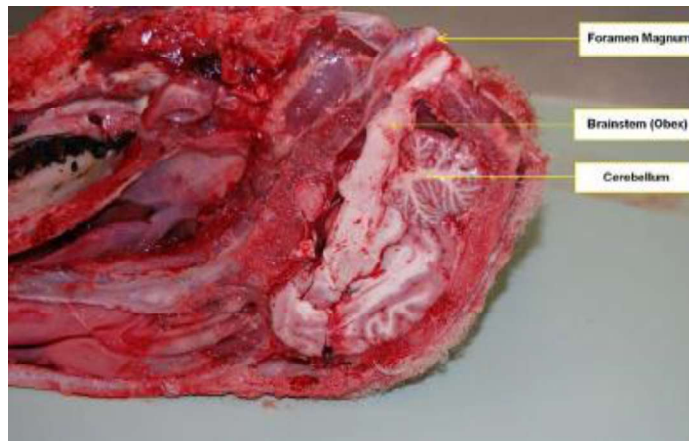


Figure 1 - Cross section of sheep head showing position of key structures involved in sample collection.



Figure 2 - Cross section of sheep head, showing the orientation of sampling spoon during first insertion. This action separates the cerebellum from the brainstem and also partially detaches the base of the brainstem.

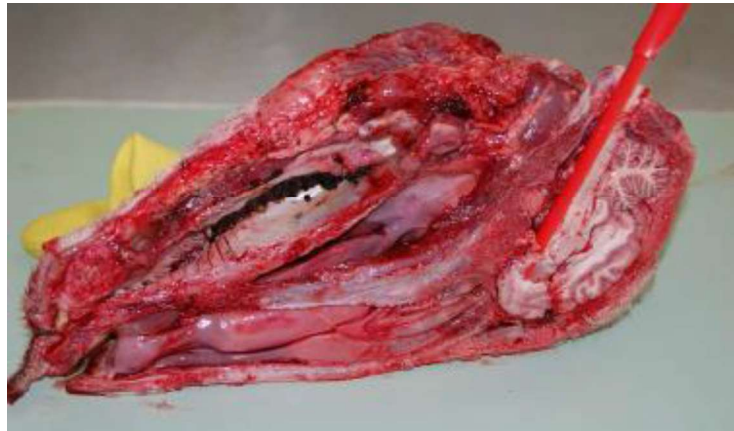


Figure 3 - Cross section of sheep head, showing the orientation of sampling spoon during second insertion. This action completes the detachment of the brainstem and allows it to be carefully withdrawn with the spoon.

2. Use the triplicate barcodes provided to label:
 - submission form (in the box marked 'Attach Barcodes')
 - sample pot
 - mini grip bag containing the ear sample and ear tag

Autolysed (Rotten) Samples

1. All brain samples can be tested despite how autolysed, or rotten, they are. If there is no structure to the brain that can be recognised, fill the pot about half full with the brain material. Ensure the cap is fully tightened to avoid any leakage.
2. If for any reason it is not possible to obtain a satisfactory sample, try the next random animal to be sampled.

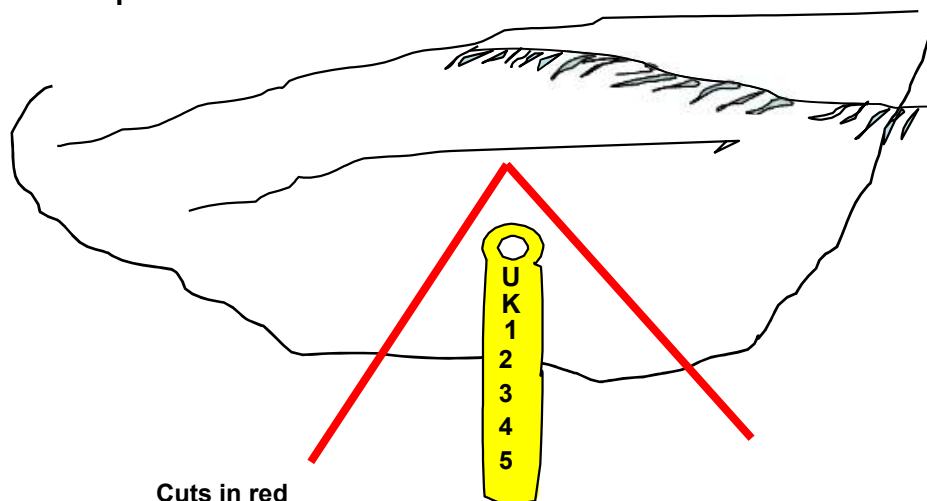
Ear Sampling

1. APHA will require a piece of ear to be removed (a 'V' shaped notch of ear surrounding the ear tag as shown in the diagram below) and placed in a clean plastic bag. If there are more than one ear tag, all tags should be included in the plastic bag and the details noted on the Submission Form (SS2). Regardless of the number of ear tags, one piece of ear sample is sufficient for DNA analysis.
2. All individual ear samples, labelled with the corresponding barcode to the sample pot and submission form, should be packaged in the same Bioshield box for transit.

3. DNA analysis can be carried out on the ear and brain tissue to ensure that the animal submitted corresponds with the sample tested and a secure trail is available should subsequent tracing and audit be required.
4. If there is any mismatch of brain material against ear sample, further action will be taken.

Diagram 4

Sheep Ear



Consumable Items

1. It is very important that consumable items not be over-ordered, wasted or confused with the Fallen Cattle Survey. The Sheep and Goat Fallen Stock Consumable Re-order Form is only to be used for this survey.

Ordering

1. Numbers of items ordered must be consistent with the numbers of samples taken at a site. All orders for this survey must be sent to EFS as detailed on the form. EFS will adjust orders as appropriate, based on known throughput of the site and the known levels of consumables previously ordered. All orders are monitored and reviewed.
2. **Refer to Appendix 12.**

Sample Pots

1. Use 100ml containers for the brain sample. Ensure the pots are not contaminated and clearly labelled with the corresponding barcode.

Spoons

1. Disposable plastic spoons must be used for all fallen stock. Metal spoons, air or water extraction methods cannot be used for sheep and goat fallen stock.

2. Spoons should not break if used correctly, apply minimal pressure. The spoons for the sheep/goat brainstem removal are RED whilst the ones used for cattle are BLUE or GREEN: the cattle spoons must not be used to sample sheep or goats.

Pots

1. A common cause of complaints from the testing laboratory is that the tops of pots have not been screwed on correctly. This can lead to leakage during transit, making the packaging difficult to handle, and also causing more expense as contaminated packaging has to be sent for specialist burning rather than being recycled.

Forceps

1. A pair of forceps is provided for each head. These must be treated as clinical waste.

Disposable Scissors

1. These should be used to cut the dura mater (membrane surrounding the brain and spinal cord), to expose the brainstem, and then used again to gently cut the cranial nerves.

Ear Sample Bags

1. EFS will supply zip top mini grip bags for storing individual ear samples. These bags should be labelled with the barcode corresponding to the brain sample and contain the ear sample and embedded ear tag of the animal. These bags can be ordered through the consumables order.

Barcode Labels

1. Each site will be supplied with rolls of barcode labels (each barcode in triplicate). A label is attached to the pot, the submission form, and the bag containing the ear sample with ear tag.

Leg Tags

1. Carcasses that are suitable for the survey will be leg tagged when collected from the farm, and sheep and goats for sampling can then be selected from leg tagged carcasses at the sampling sites.
2. Leg tags can be ordered by using a consumables order form. These are exactly the same as the ones used for the fallen cattle survey but ensure the correct number is ordered through appropriate survey re-order forms. The leg tag number would be matched to the collectors' commercial documentation to verify ID. If owners take their own fallen sheep or goats to the plants, leg tagging and selection can be done at the plants.

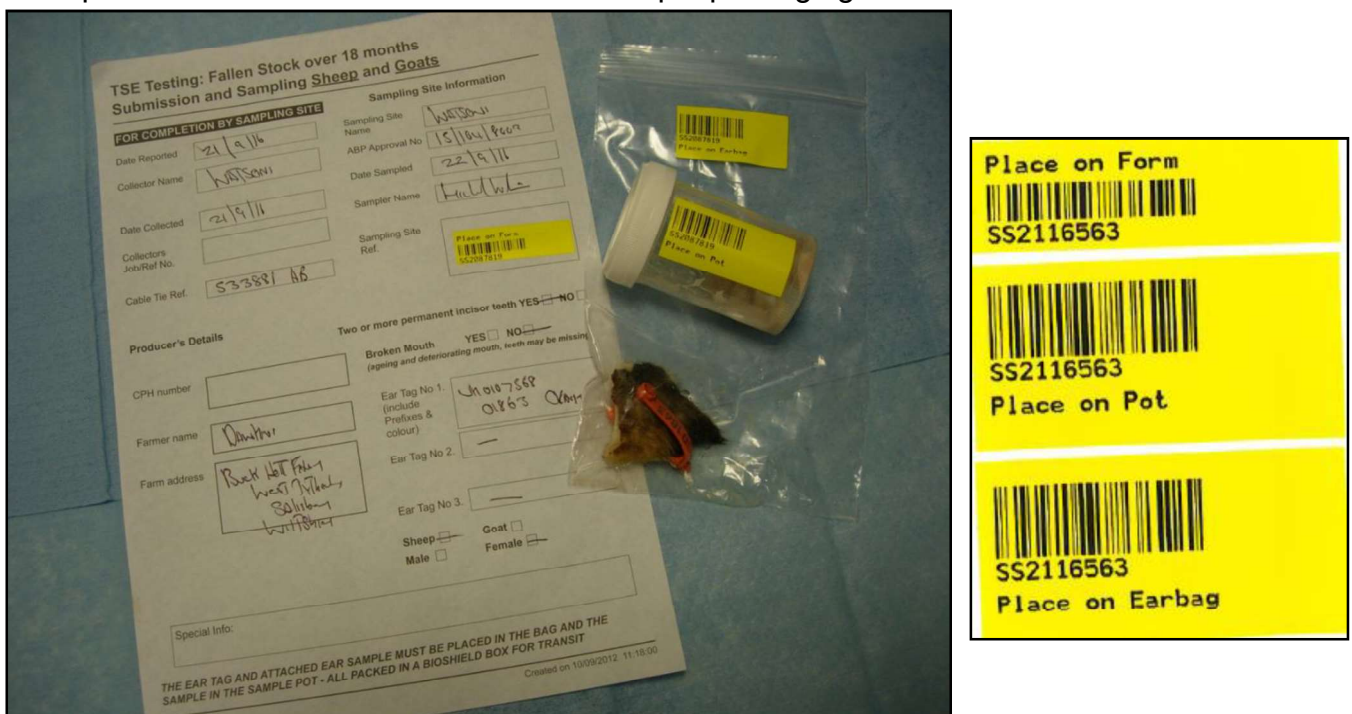
Packaging

1. Place the brainstem sample and cerebellum in the 100ml sample pot:

- Ensure that the lid is securely on the sample pot to avoid leakage of autolysed sample or blood.
- Label the pot with a barcode from the triplicate yellow barcodes provided ('Place on Pot').
- Place the eartag, with a piece of ear, in the ear bag labelled with a yellow barcode from the triplicate barcodes ('Place on Earbag') so that the ear and pot have the same unique barcode number.
- Ensure that the ear bag is sealed.
- Place the sample pot in the Bioshield box with the ear bag on top and seal well.
- Place a fully completed sample submission form for each sample (SS2) inside the Bioshield box.
- The operator should ensure that the remaining yellow label from the triplicate barcodes received is placed onto this corresponding submission form ('Place on Form').
- Place all sample submission forms inside a sealable plastic bag and place the bag with the forms into the box with the samples and ear bags.
- Seal the Bioshield box tightly.

Figure 4

Sheep Fallen Stock submission form and sample packaging.



3. Currently all packaging needs to be requested through EFS.
4. Courier Topspeed contact numbers are 01565 631 840 or 0800 856 2464 and quote FALLEN_STOCK.
5. Cattle, sheep and goat sample pots can be sent in the same box, if necessary, to reduce packaging.
6. There are three packaging containers available under this survey:

- Small Bioshield Box - (up to six samples)
 - Medium Bioshield Box - (up to ten samples)
 - Large Bioshield Box - (up to 40 samples)
7. Ensure all packaging is available before sampling commences and samples are packed as instructed and stored correctly until collection.

Despatch of Samples

1. Each site has a responsibility to notify EFS as to the number of samples being sent.
2. Complete the attached form and fax EFS at 01925 248876 or email to bse.services@forensicsuk.eurofins.com.
3. **Refer to Appendix 8.**
4. The sampling site is responsible for having the sample box packed correctly and ready for collection by the courier.
5. **Refer to Appendix 7.**
6. Sites sampling on a less frequent basis may have a limited collection service. In these circumstances, samples must be stored in a refrigerator before collection. Samples should not be stored for more than three days. Samples sent on the same day of sampling will yield much better test results rather than stored samples and therefore we ask that the sites co-operate with the laboratories to get the best outcome.
7. APHA monitor the condition of packaging and all sites will be encouraged to keep packaging clean, pack in accordance with instructions and reduce contamination to a minimal level within the Bioshield bag. As stated previously boxes can only be recycled if they are not contaminated.

Payment for Sampling

1. Sampling payments will be organised monthly and based on the number of samples tested at APHA by the last day of the calendar month.
2. Due to the time taken for both delivery and testing, some sampling will go across to the following month's payment.
3. APHA will send an invoice request to the sampling sites usually in the second week of the month. The invoice would show the samples submitted by individual sites. The sampling site would be required to create an invoice and send it by email (APInvoices-APH-U@gov.sscl.com) or post to Shared Services Connected Limited, DEF Procure to Pay, PO Box 790, Newport, Gwent, NP10 8FZ. The APHA will pay the provider

meeting the requirements stipulated £10.00 (+ VAT) per sample. This was increased from the 1st January 2026, from £7.25(+ VAT). For any issues relating to payment for sampling, contact 020 8415 2216.

4. The Cattle sampling fee will no longer be paid as of 1 April 2019, except under the circumstances outlined above for approved sampling sites, but there are no changes to sheep and goat sampling fee or the process of submitting the invoice.

Further Information

1. We would encourage all sampling sites to look at relevant pages on the APHA website for further information.

Additional Information CSFS

1. For every carcase to be collected, an NSP60 form will be attached to the notification email. Adhere to the following instructions:
 - The NSP60 form that confirms the animal's ID should accompany the carcase, and will need to be partially completed by you (section headed For Completion at Sampling Premises). The form must give the CPH that the animal has been submitted by.
 - Additional SS2 forms or labels must **NOT** be used for CSFS fallen stock under any circumstances.
 - Do not add any barcode labels to the NSP60 or the sample pot that accompanies the NSP60. Instead, write 'CSFS' on the sample pot.
2. When the above carcasses have been collected, brainstem samples will need to be taken and sent to EFS for TSE testing.
3. Place the brainstem sample and cerebellum in the 100ml sample pot:
 - Ensure that the lid is secure on the sample pot to avoid leakage of autolysed sample or blood.
 - Label the sample pot with a permanent black marker/pen with a letter or number, e.g. 'A' or '1'.
 - Place the eartag, with a piece of ear, in the ear bag labelled with the same number or letter that was used for the corresponding sample pot, e.g. 'A' or '1'.
 - Ensure that the ear bag is sealed.
 - Place the sample pot in the Bioshield box with the ear bag on top and seal well.
 - Place a fully completed sample submission form for each sample (NSP60) inside the Bioshield box.
 - Label the submission form with the same number or letter that was used for the corresponding sample pot and ear bag, e.g. 'A' or '1' for efficient identification at the laboratory.
 - Place all sample submission forms inside a sealable plastic bag and place the bag with the forms into the box with the samples and ear bags.
 - Seal the Bioshield box tightly.

6. Fallen stock centre must ensure that this sample is not charged to APHA.
7. Sampling sites can use the existing APHA courier service, but not solely for the transport of SMS / SQS samples. Keep the SMS / SQS samples in a refrigerator until they can be added to the next APHA courier collection. (If that is not due in the following week contact the Premium Sheep and Goat Health Scheme (PSGHS) office for guidance (SRUC) 01835 822456).
8. NFSCo will pay the fallen stock collector the previously agreed fee for SMS / SQS collection and sampling.
9. Any queries, contact NFSCo and they will contact SRUC Veterinary Services.
10. If correct process is NOT followed and the test outcome is incorrectly reported, the farmer will not be able to comply to the schemes.

Appendix 1: Ear tag Country Code

Country of birth	Ear tag country code
Austria	AT
Belgium	BE
Bulgaria	BG
Croatia	HR
Cyprus	CY
Czech Republic	CZ
Denmark	DK
Estonia	EE
Finland	FI
France	FR
Germany	DE
Greece	EL
Hungary	HU
Ireland (ROI)	IE
Italy	IT
Latvia	LV
Lithuania	LT
Luxembourg	LU
Malta	MT
Netherlands	NL
Poland	PL
Portugal	PT
Romania	RO
Slovakia	SK
Slovenia	SI
Spain	ES
Sweden	SE
UK (including Channel Islands and Isle of Man)	UK
All other countries	UK (unless slaughtered within 20 days imported. Import information is shown on the inside back page of the cheque book style passport)

Appendix 2: FSCA 2 Submission Form

Bovine Carcase Collection and Sample Submission Form for TSE Surveillance of Fallen Cattle Aged Over 48 Months

Please select material type **Category 1** – For Disposal Only **Category 2** – Not For Animal Consumption

FOR COMPLETION BY COLLECTOR	
Collectors Job or Ref No.	<input type="text"/>
Time and Date of Notification	<input type="text"/>
Cable Tie Ref.	<input type="text"/>
Ear Tag No.	<input type="text"/>
Date of Birth	<input type="text"/>
PRODUCER'S DETAILS	
CPH No.	<input type="text"/>
Name	<input type="text"/>
Address <i>If carcase not at this address, please give details in Special Info Section</i>	<input type="text"/>
Movement Card Attached	Yes <input type="checkbox"/> No <input type="checkbox"/>
Animal Alive	Yes <input type="checkbox"/> No <input type="checkbox"/>
Sex of animal	M <input type="checkbox"/> F <input type="checkbox"/>
Clinical History (must tick a box as appropriate)	
Changes in behaviour	Yes <input type="checkbox"/> No <input type="checkbox"/> NK <input type="checkbox"/>
Sensitive to touch/sound	Yes <input type="checkbox"/> No <input type="checkbox"/> NK <input type="checkbox"/>
Uncoordinated	Yes <input type="checkbox"/> No <input type="checkbox"/> NK <input type="checkbox"/>
Other Neurological Signs	Yes <input type="checkbox"/> No <input type="checkbox"/> NK <input type="checkbox"/>
<i>(NK* – not known)</i>	
Time and Date of Death	<input type="text"/>
Reason for Death <i>if known</i>	<input type="text"/>
Tel. No.	<input type="text"/>
Special Information (e.g. VICs Submission ID)	<input type="text"/>
COLLECTOR DETAILS	
Company Name	<input type="text"/>
Company Address	<input type="text"/>
Company Tel. No.	<input type="text"/>
Time and Date of Collection	<input type="text"/>
Driver Name and Signature	<input type="text"/>
FOR COMPLETION AT SAMPLING SITE	
Sampling Site Name	<input type="text"/>
ABP Approval No.	<input type="text"/>
Sampling Site Address	<input type="text"/>
Tel. No.	<input type="text"/>
Hide ID No. <i>if applicable</i>	<input type="text"/>
Sampling Site Reference No.	ATTACH BARCODE LABEL HERE
Date and Time of Sample Collection	<input type="text"/>
If Not Sampled, Give Reason	<input type="text"/>
Sampler Name and Signature	<input type="text"/>
Date Sent to Lab	<input type="text"/>

FSCA2 Ver.1 (Rev 05/24)

https://cdnmedia.eurofins.com/european-west/media/u0nfgqj/fsc2-ver-1-rev-12-08_final_1july2024.pdf

Appendix 3: Cascade Training Form



BRAINSTEM REMOVAL CASCADE TRAINING FORM

NOTIFICATION OF NAMES OF STAFF TRAINED IN REMOVAL OF BRAINSTEM FROM : * CATTLE SHEEP & GOAT

*Please tick species to which training applies

Plant / Site Name : _____

Plant / Site Number : _____
(if applicable)

Plant / Site Tel Number : _____

Plant / Site Fax Number : _____

FSA Region Number : _____
(if applicable)

Persons Trained:-

Name	Age	Date(s) of Training	Trainer

Name(s) of Staff Originally Trained or Assessed by APHA : _____

Name of original APHA Trainer or assessor : _____

Signed : _____

Name in capitals : _____

Position (e.g. plant manager) : _____

Date : _____

Submit completed form to :
 DSG, TSE Team, 2nd Floor, Area E, Weybourne Building, Woodham Lane, New Haw, Addlestone, Surrey KT15 3NB
 or email it to : tse.queries@apha.gov.uk

Cascade Training Form – Ver9 (Oct 23)

Appendix 4: Sample Assessment Form



Sample Assessment Submission Form

This assessment is only suitable for operators who have previously received training in brainstem removal.

Instructions :

Each operator must select **SIX** fresh under 48 month animals. Remove brainstem samples using the standard disposable spoon method. Decomposed samples must **not** be submitted. Place the suitable samples into a specimen pot and label with the UK ear tag number. Pack the specimen pots in a Biosheild box, together with this form, and despatch to Eurofins Forensic Services Ltd (EFS), **Risley**. Use one form per operator. Please mark the outside of the box with "**Assessment Only – Not For Test**".

Name of Site:	
Name of Operator:	
Originally Trained By :	
Original Training Date :	
When did you last routinely collect samples ?	

Date Assessment samples taken :			
Sample	UK Tag Number	Condition Score <i>Lab use only</i>	Assessed By <i>Lab use only</i>
1			
2			
3			
4			
5			
6			

Comments :

The Samples submitted give / do not give* sufficient evidence to demonstrate operator competence in brainstem removal technique.

* Delete as appropriate

Authorised by :	Date
On behalf of EFS Risley	

Lab must return completed form to :

DSG, TSE Team, 2nd Floor, Area E, Weybourne Building, Woodham Lane, New Haw, Addlestone, Surrey KT15 3NB or email it to : tse.queries@apha.gov.uk

Ver6 – January 21

Appendix 5: Bovine Consumable Order Form



Forensic Services

Bovine Fallen Stock (TS5902) Consumable Re-Order Form

Site Name:	
Address:	
Post Code:	Telephone:
Contact Name:	Sign:

Please use this form to order items

Description	Item Code	Unit of Issue	Units required	EFS Use Only
Forms	LGC031	Pad of 50		
Leg Cable Ties	LGC028	Pack of 100		
Disposable Spoon	LGC032	Pack of 20	<input type="text"/>	
Disposable forceps	LGC006	Pack of 20	<input type="text"/>	
90ml sample pots	LGC004	Pack of 100	<input type="text"/>	
Ear sample bags	LGC017	Pack of 100		
Barcode Stickers	—	Roll of 1000		
Bio - Box (Medium 6 samples)	LGC035	Each		
Bio - Box (Large 40 samples, Box only)	LGC036	Each		
Techni-Ice Sheet	LGC018	Pack of 10		

Please fax to: **01925 248876**

OR Post to:

**TSE Department
Eurofins Forensic Services
Darwin House
Faraday Street
Birchwood Park
Risley
WA3 6FW**

OR Email to:
bse.services@forensicsuk.eurofins.com

For EFS use only

Goods Approved for dispatch

By:

Date:

Appendix 6: Consumable Guidance

Consumables Order Guidance

This table is only to be used as a guide. Your own stock levels must be regularly checked and consumable requests should be based on these checks.

Suggested Consumables Item request	Expected Sample Throughput				
	Less than 10 per week	10 – 25 per week	25 – 50 per week	10 – 50 per day	More than 50 per day
Form FSCA2 x50	4	10	20	40	80
Leg Cable Ties	200	500	1000	2000	4000
Spoons x20	10	25	50	100	200
Sample Pots x250	1	2	4	8	16
Ear sample bags x100	2	5	10	20	40
Disposable forceps x500	1	1	2	4	8
Barcode stockers x2500	1 Roll	1 Roll	1 Roll	2 Rolls	2 Rolls
Bio-Box Small x20	2	3	0	0	0
Bio-Box Medium x1	40	100	100	0	0
Bio-Box Large x1	0	5	25	50	100
Tech-Ice Sheet x1	50	100	125	50	100
Suggested Re-Order Level Based on pots & spoons	50	100	200	500	1000

Last Update: Oct/23

Appendix 7: Box Packing Guide

Bioshield Box Packaging Guide – Prepared by AHVLA Newcastle



1) Prepare the Techni-Ice sheet, as instructed on the sheet, the day before despatch of samples.



2) Ensure all sample container lids are tightly screwed shut and exterior of containers are wiped clean. Place samples in the rack.



3) Place the rack & containers into the Pathoseal bag. Force excess air from the bag, remove the peelable tab and fully seal the bag shut.



4) The two polystyrene sheets should be placed in the bottom of the outer shipping box.



5) Put the sealed tray of sample containers into the outer shipping box.



6) The frozen pre-prepared Techni-Ice sheet now be placed on top of the sealed sample tray.



7) Place all relevant documents in the supplied document bag and put these on top of the Techni-Ice sheet.



8) Close the box, attach a completed Hays tracking label and seal the box with the blue security sticker.

Important Note

Boxes showing any signs of external contamination, with blood or tissue, must **NOT** be despatched. If this happens, Repack the samples in a new outer shipping box before despatch.

AHVLA Newcastle – July 2014 v2

Appendix 8: Notification of Samples Sent Form



NOTIFICATION OF TSE SAMPLES SENT TO EFS FOR TESTING

SAMPLING SITE
SAMPLING DATE

Sample Type	Number Sent		EFS USE ONLY
	Boxes	Samples	Submission Received as expected (Y/N) <i>If N record reason in comment box</i>
Cattle			
Sheep			
Goat			
Other			

Transport Type:

Please Tick Topspeed	<input type="checkbox"/>	Please Tick Royal Mail	<input type="checkbox"/>	Please Tick Other	<input type="checkbox"/>
--------------------------------	--------------------------	----------------------------------	--------------------------	-----------------------------	--------------------------

Sender's Signature

Sender's Name
(BLOCK CAPITALS)

Date Sent

Comments :

PLEASE EMAIL ON DAY OF SAMPLE DESPATCH TO
bse.services@forensicsuk.eurofins.com

OR ALTERNATIVELY FAX ON THE DAY OF DESPATCH TO
 01925 248876

Sample Submission Notification V4 September 2023

https://cdnmedia.eurofins.com/european-west/media/19028291/efs-sample-submission-notification-ver4_sept-2023.pdf

Appendix 9: Cattle Brainstem Sample Quality Guide

Cattle Brainstem Sampling

Upper surface view



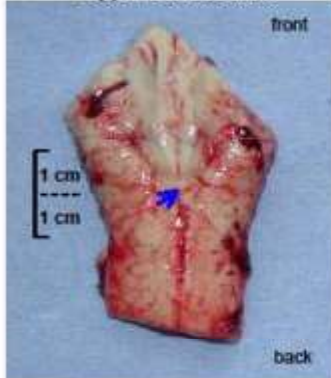
An Ideal sample

With care and practice, the operator should be able to consistently produce samples with 2 cms or more undamaged brainstem both in front and behind the point of the 'V' shape visible on the upper surface (arrowed)

Lower surface view



Upper surface view




A Marginal sample

The minimum required to ensure the sample will be suitable for testing is 1 cm of undamaged brainstem both in front and behind the point of the 'V' shape visible on the upper surface.

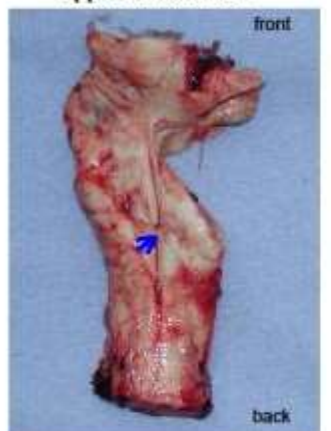
This sample is adequate, but only just !

Loss of tissue or damage at the back of the sample is usually due to careless removal of the head. Loss or damage to the front of the sample is due to poor sampling technique.

Lower surface view



Upper surface view




A Poor Sample

Damage during sampling; less than 1 cm undamaged brainstem in front of the point of the 'V' shape.

Such samples should still be sent to the laboratory, but may prove unsuitable for testing.

THE LABORATORY WILL DO ITS BEST TO TEST UNSATISFACTORY SAMPLES, BUT IF THIS PROVES IMPOSSIBLE, THE CARCASE WILL NOT BE RELEASED FOR HUMAN CONSUMPTION AND NO COMPENSATION WILL BE PAID.

Lower surface view



Appendix 10: Sheep and Goat Sample Submission Form (SS2)

TSE Testing: Fallen Stock over 18 months Submission and Sampling Sheep and Goats

FOR COMPLETION BY SAMPLING SITE

Date Reported

Collector Name

Date Collected

Collectors Job/Ref No.

Cable Tie Ref.

Sampling Site Information

Sampling Site Name

ABP Approval No

Date Sampled

Sampler Name

Sampling Site Ref.

Producer's Details

Two or more permanent incisor teeth YES NO

CPH number

Farmer name

Farm address

Broken Mouth YES NO
(ageing and deteriorating mouth, teeth may be missing)

Ear Tag No 1. (include Prefixes & colour)

Ear Tag No 2.

Ear Tag No 3.

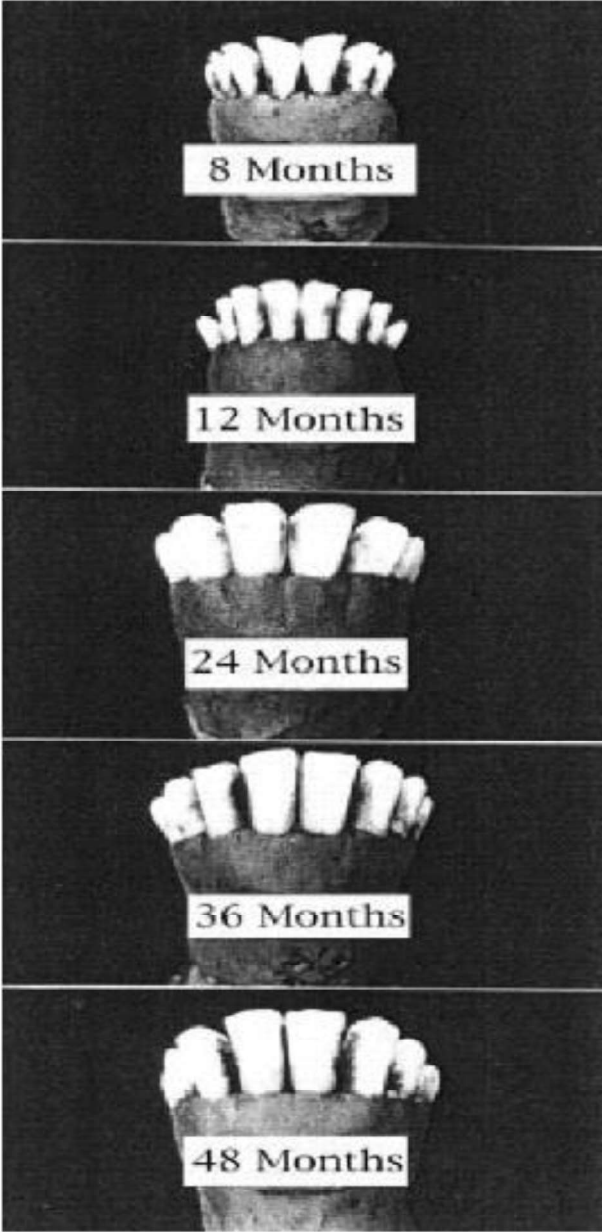
Sheep Goat
Male Female

Special Info:

THE EAR TAG AND ATTACHED EAR SAMPLE MUST BE PLACED IN THE BAG AND THE SAMPLE IN THE SAMPLE POT - ALL PACKED IN A BIOSHIELD BOX FOR TRANSIT

Appendix 11: Incisor Teeth Guide

Sheep & Goat Teeth
Guide to identifying temporary and permanent incisor teeth of sheep and goats.



Appendix 12: Sheep and Goat Consumables Re-order Form

**SHEEP & GOAT SCRAPIE SURVEY
(TS-5902)
CONSUMABLES RE-ORDER FORM (V4)**



Forensic Services

Plant Name:.....	Please fax or email to: 01925 248876 bse.services@forensicsuk.eurofins.com TSE Laboratory Eurofins Forensic Services Darwin House Faraday Street Birchwood Park Risley WA3 6FW
Address.....	
..... Post Code.....	
Telephone.....	
Contact Name.....	

Please complete stating how many of each you require

Description	Item Code	Unit of Issue	Quantity Required	EFS Use only
Bioshield Box 40 Pot Tray (No pots)	LGC001	Each		
Bioshield Box 6 Pot Tray (No pots)	LGC037	Each		
Bioshield Box 10 Pot Tray (No pots)	LGC038	Each		
100 ml containers	LGC004	Pack of 100		
Red Ear Tags <i>(only applicable for Abattoirs)</i>	LGC005	Strip of 20		
Forceps	LGC006	Box 20		
Disposable Scissors	LGC007	Box 25		
Red Spoons	LGC008	Pack of 100		
Scalpels No.11	LGC009	Pack of 10		
Benchkote	LGC010	Pack 100 sheets		
Spray Paint Green	LGC011	Aerosol Each		
Masks (FFP2)	LGC012	Pack of 10		
Goggles	LGC013	Pair		
Vinyl Gloves Large	LGC014	Box 100		
Vinyl Gloves Medium	LGC015	Box 100		
Vinyl Gloves Small	LGC016	Box 100		
Ziptop Mini-Grip Bags	LGC017	Pack of 100		
Techni-Ice Sheet	LGC018	Pack of 10		
Sharps Box	LGC020	Each		
Document Bags	LGC021	Pack of 100		
Blue Paper	LGC022	Roll		
Disposable Aprons	LGC023	Pack of 200		
BioSeal Leakproof Bag	LGC024	Pack of 10 (max 1pk per order)		
Black Sacks	LGC025	Pack of 100		
Black Marker Pen	LGC026	Each		
Cable Ties	LGC028	Pack of 100		
Barcode Stickers	LGC029	Roll of 200		
Submission Forms	LGC030	50 sets of forms per Pad		

https://cdnmedia.eurofins.com/european-west/media/21342179/sheep-and-goat-re-order-ver4_sept-2023.pdf

Appendix 13: FSCA 2 Submission Form for Cohorts

COHORT

Bovine Carcase Collection and Sample Submission Form for TSE Surveillance of Fallen Cattle Aged Over 48 Months

Please select material type Category 1 – For Disposal Only Category 2 – Not For Animal Consumption

FOR COMPLETION BY COLLECTOR

Collectors Job or Ref No. <input style="width: 95%;" type="text"/> Time and Date of Notification <input style="width: 95%;" type="text"/>	Cable Tie Ref. <input style="width: 95%;" type="text"/> Ear Tag No. <input style="width: 95%;" type="text"/> Date of Birth <input style="width: 95%;" type="text"/>
--	---

PRODUCER'S DETAILS

CPH No. <input style="width: 95%;" type="text"/> Name <input style="width: 95%;" type="text"/> Address <input style="width: 95%; height: 40px;" type="text"/> <small><i>If carcase not at this address, please give details in Special Info Section</i></small> Tel. No. <input style="width: 95%;" type="text"/>	Movement Card Attached Yes <input type="checkbox"/> No <input type="checkbox"/> Animal Alive Yes <input type="checkbox"/> No <input type="checkbox"/> Sex of animal M <input type="checkbox"/> F <input type="checkbox"/> Clinical History (must tick a box as appropriate) Changes in behaviour Yes <input type="checkbox"/> No <input type="checkbox"/> NK <input type="checkbox"/> Sensitive to touch/sound Yes <input type="checkbox"/> No <input type="checkbox"/> NK <input type="checkbox"/> Uncoordinated Yes <input type="checkbox"/> No <input type="checkbox"/> NK <input type="checkbox"/> Other Neurological Signs Yes <input type="checkbox"/> No <input type="checkbox"/> NK <input type="checkbox"/> <small>(NK* – not known)</small> Time and Date of Death <input style="width: 95%;" type="text"/> Reason for Death <i>if known</i> <input style="width: 95%;" type="text"/> Special Information (e.g. VICs Submission ID) <input style="width: 95%;" type="text"/>
---	--

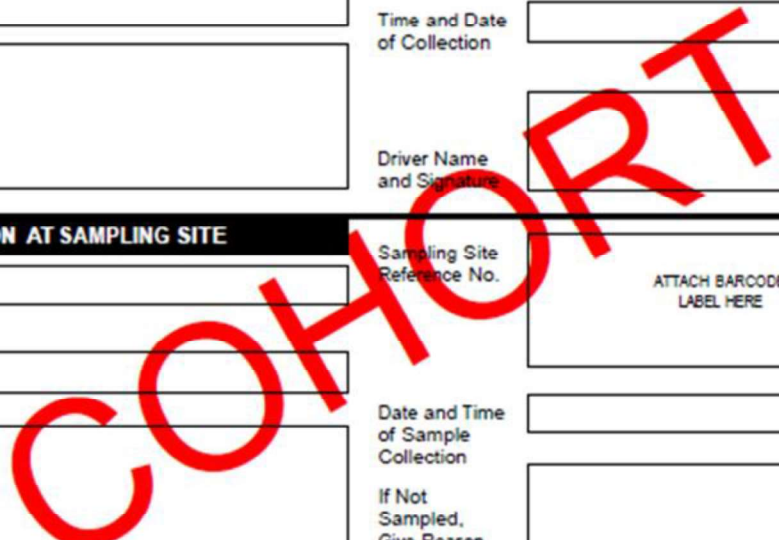
COLLECTOR DETAILS

Company Name <input style="width: 95%;" type="text"/> Company Address <input style="width: 95%; height: 40px;" type="text"/>	Company Tel. No. <input style="width: 95%;" type="text"/> Time and Date of Collection <input style="width: 95%;" type="text"/> Driver Name and Signature <input style="width: 95%; height: 30px;" type="text"/>
---	---

FOR COMPLETION AT SAMPLING SITE

Sampling Site Name <input style="width: 95%;" type="text"/> ABP Approval No. <input style="width: 95%;" type="text"/> Sampling Site Address <input style="width: 95%; height: 40px;" type="text"/> Tel. No. <input style="width: 95%;" type="text"/> Hide ID No. <i>if applicable</i> <input style="width: 95%;" type="text"/>	Sampling Site Reference No. <input style="width: 95%;" type="text"/> Date and Time of Sample Collection <input style="width: 95%;" type="text"/> If Not Sampled, Give Reason <input style="width: 95%; height: 30px;" type="text"/> Sampler Name and Signature <input style="width: 95%; height: 30px;" type="text"/> Date Sent to Lab <input style="width: 95%;" type="text"/>
--	---

FSCA2 Ver.1 (Rev 05/24)



Appendix 14: FSCA 2 Submission Form for Homekill

HOME KILL

Bovine Carcase Collection and Sample Submission Form for TSE Surveillance of Fallen Cattle Aged Over 48 Months

Please select material type Category 1 – For Disposal Only Category 2 – Not For Animal Consumption

FOR COMPLETION BY COLLECTOR	
Collectors Job or Ref No.	Cable Tie Ref.
Time and Date of Notification	Ear Tag No.
	Date of Birth
PRODUCER'S DETAILS	
CPH No.	Movement Card Attached Yes <input type="checkbox"/> No <input type="checkbox"/>
Name	Animal Alive Yes <input type="checkbox"/> No <input type="checkbox"/>
Address <i>If carcase not at this address, please give details in Special Info Section</i>	Sex of animal M <input type="checkbox"/> F <input type="checkbox"/>
	Clinical History (must tick a box as appropriate)
	Changes in behaviour Yes <input type="checkbox"/> No <input type="checkbox"/> NK <input type="checkbox"/>
	Sensitive to touch/sound Yes <input type="checkbox"/> No <input type="checkbox"/> NK <input type="checkbox"/>
	Uncoordinated Yes <input type="checkbox"/> No <input type="checkbox"/> NK <input type="checkbox"/>
	Other Neurological Signs Yes <input type="checkbox"/> No <input type="checkbox"/> NK <input type="checkbox"/>
	(NK* – not known)
	Time and Date of Death
	Reason for Death <i>If known</i>
Tel. No.	Special Information (e.g. VICs Submission ID)

COLLECTOR DETAILS	
Company Name	Company Tel. No.
Company Address	Time and Date of Collection
	Driver Name and Signature

FOR COMPLETION AT SAMPLING SITE	
Sampling Site Name	Sampling Site Reference No.
ABP Approval No.	ATTACH BARCODE LABEL HERE
Sampling Site Address	
	Date and Time of Sample Collection
	If Not Sampled, Give Reason
Tel. No.	Sampler Name and Signature
Hide ID No. <i>If applicable</i>	Date Sent to Lab

FSCA2 Ver.1 (Rev 05/24)



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Any enquiries regarding this publication should be sent to us at Email tse.queries@apha.gov.uk

Using and sharing your information

How we use your personal data is set out in our personal information charter and privacy notices:

- <https://www.gov.uk/government/organisations/department-for-environment-food-rural-affairs/about/personal-information-charter>
- <https://www.gov.uk/government/publications/animal-and-plant-health-agency-privacy-notice>

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